

R. E. Gaensslen,<sup>1</sup> Ph.D.; Suzanne C. Bell,<sup>2</sup> M.S.; and Henry C. Lee,<sup>3</sup> Ph.D.

## Distributions of Genetic Markers in United States Populations: III. Serum Group Systems and Hemoglobin Variants

---

**REFERENCE:** Gaensslen, R. E., Bell, S. C., and Lee, H. C., "Distributions of Genetic Markers in United States Populations: III. Serum Group Systems and Hemoglobin Variants," *Journal of Forensic Sciences*, JFSCA, Vol. 32, No. 6, Nov. 1987, pp. 1754-1774.

**ABSTRACT:** All published and unpublished population frequency data that could be located for U.S. populations are tabulated and presented for the serum group systems haptoglobin ( $\alpha$ -chain), group specific component, and transferrin and for the common  $\beta$ -chain variants of hemoglobin. Results obtained by combining data for comparable racial/ethnic groups are also presented. Some evidence is presented to indicate that the results obtained from the combined data may give better information on frequencies for the U.S. population at large than is obtainable from studies conducted in restricted geographic areas.

**KEYWORDS:** forensic sciences, genetic typing, demography, population genetics, United States populations, genetic markers, genotypic frequencies, phenotypic frequencies, isoenzyme systems, human serum protein polymorphism, haptoglobin system, group specific component system, vitamin D binding protein, transferrin system, hemoglobin  $\beta$ -chain variants

The growth and development in forensic serology in the past 25 years has revealed a substantial number of genetic marker systems from which routine parentage testing protocols may be constructed [1-3] and for the partial individualization of blood and physiological fluid stains [4-10]. Interpretation of the significance of typing results in criminalistics applications and calculations of the probability of paternity in nonexclusion parentage cases both require knowledge of genotypic and phenotypic frequencies in applicable populations.

Thousands of frequency studies on various genetic marker systems have been carried out on many populations throughout the world, the most complete compilation of them being the extraordinary work by Mourant et al. [11]. World population data on the haptoglobin (HP) and group specific component (GC) systems were compiled and summarized in 1969 [12]. The most complete compilation of population studies on hemoglobin variants was published by Livingstone in 1967 [13]. Genetic marker frequency data from the many different

Various aspects of this work were presented in part at the 8th Annual Meeting of the Northeastern Association of Forensic Scientists, Albany, NY, 1982; the 35th Annual Meeting of the American Academy of Forensic Sciences, Cincinnati, OH, 15-19 Feb. 1983; and the International Symposium on Forensic Applications of Electrophoresis, FBI Academy, Quantico, VA, 1984. Received for publication 5 Jan. 1987; accepted for publication 11 Feb. 1987.

<sup>1</sup>Professor and director, Forensic Sciences Program and Laboratories, University of New Haven, West Haven, CT.

<sup>2</sup>Research chemist, Los Alamos National Laboratories, Los Alamos, NM.

<sup>3</sup>Chief criminalist and director, Connecticut State Police Forensic Science Laboratory, Meriden, CT.

studies of U.S. populations, however, have not to our knowledge been thoroughly compiled. In this paper, we summarize all the published and some unpublished population frequency data that could be located for U.S. populations for the three serum group systems haptoglobin (HP), group specific component (vitamin D binding protein; GC), and transferrin (TF), and for the common  $\beta$ -chain variants of hemoglobin (HBB), along with some results obtained by combining data from different studies. This paper along with two previous companion ones [14, 15] provides a summary and analysis of U.S. population data for 22 genetic marker systems.

## Methods

The conventions used in presenting the tabular data as well as the methods used in combining comparable data from different population studies were fully described previously [14, 15]. Briefly, within the separate tables, each representing a different system, data are tabulated separately for Caucasian, Negro, Hispanic, Asian, or other populations. Each population studied is identified by location (two-letter abbreviations are used for states) and by a description of the blood donors if provided. A reference is given for each study. References to population studies are given a "T" (for "Table") prefix in the tables and are separately compiled at the end of the paper. Data for each phenotype within each system are reported using an  $NNN(\%. \%)$  format, where  $NNN$  represents the number of individuals who possessed the phenotype and  $\%. \%$  represents the percentage rounded to one decimal place. The total number of people studied is also given, and is not always the sum of the major phenotypes because rare phenotypes were observed. The notes in each table provide data for subtypes, rarer types, unusual or descriptive features of a population, or explanations about the calculations. Data from GC\*1 and TF\*C subtype studies were used to compute "electrophoretic" GC and TF phenotype frequencies in cases where this was not done in the original paper.

Two calculations were used to combine all the data for a particular racial/ethnic class within a genetic marker system, where a sufficient number of different studies were available for comparatively similar groups within that class.

The first sums the numbers of individuals for all data sets showing numbers, and a percentage value for each phenotype is computed from the resulting totals, yielding what is referred to as the "numerical total." The second weights the percentage distributions (proportions) for each phenotype according to the number of individuals typed and yields what is referred to as the "weighted mean of proportions," or "WMP." A weighted standard deviation of proportions (WSDP) was also calculated for each WMP. Details were given in the previous papers [14, 15]. Data reported in numbers of individuals were used to calculate proportions. No attempt was made to calculate numbers of individuals from data reported in percentage distributions. Hence, all data are used in computing WMP, whereas only data reported in numbers of individuals are included in the numerical total.

All available data were included in the tables for completeness, but a data set was not used in the calculations in cases where data were incomplete or authors provided only a gene frequency result.

Gene frequencies were calculated by gene counting for data sets in which there was sufficient information to enable the calculation. In HP system data, 2-1, 2-1M, and 0 types are shown separately for those studies in which they were so reported. The 2-1 and 2-1M types were combined and the HP 0 excluded from consideration, however, for calculation of gene frequencies. No separate  $HPA*2M$  frequency was calculated in these studies. The HP 0 phenotype individuals were excluded from gene frequency calculations because the genetic basis of HP 0 is not very clear. Individuals who are HP 0 are often  $HPA*2M/HPA*2M$ ,  $HPA*2M/HPA*1$ , or  $HPA*2M/HPA*2$ , but can possess  $HPA*2/HPA*1$  or  $HPA*2/HPA*2$  genotypes as well [16, 17]. In TF system data, variant TF\*B and TF\*D types ob-

served are given in the notes. TF\*B and TF\*D types were combined for gene frequency calculations, however.

A chi-square value was calculated for every data set for which gene frequencies could be meaningfully calculated and for the corresponding numerical totals. In Tables 1 and 2, data sets having  $\chi^2$  values corresponding to  $0.01 < P < 0.05$  or  $P < 0.01$  are indicated. Interpretation of chi-square values calculated for TF and HB data are complicated by the small numbers and correspondingly low gene frequencies of variants. In a number of cases, the number of phenotypic classes is the same as the number of genes whose frequencies are being estimated. In a significant number of cases, it was clear that the  $\chi^2$  values obtained were not providing meaningful indications of goodness of fit. Accordingly, gene frequencies for TF and HBB genes are reported as calculated. In the hemoglobin system, rarer phenotypes involving genes being estimated, such as HBB SC, were counted in computing gene frequencies.

The gene, genotypic, and phenotypic nomenclature rules suggested by Shows et al. [18] have been followed for all the systems. Using these rules, hemoglobin variant genes are designated according to the chain in which the substitution occurs, the position number in that chain, and the variant amino acid using one letter abbreviations. The HB sickle gene, for example, is designated HBB\*6V, indicating that this gene gives rise to a valine substitution at Position 6 in the  $\beta$  chain. Similarly, the HB C and HB D genes are designated HBB\*6K and HBB\*12IQ, respectively. The nomenclature of variant GC genes and phenotypes follows Constans and Cleve [19].

Most computations were carried out on a Data General MV 8000 mainframe computer with programs written in FORTRAN.

## Results and Discussion

Table 1 gives phenotypic and genotypic frequencies for the HP system. HP 2-1M and HP 0 types are seen in significant frequencies only in the Negro populations. The majority of the data sets including the numerical totals show good fit to Hardy Weinberg expectations on the basis of chi-square computations.

Table 2 gives the data for the GC system. As with the HP data, most of the studies exhibit goodness of fit to expectation. Table 3 gives data for the TF system. TF BC occurs more commonly in Caucasians than in Negroes, while TF CD is seen in some 5% of Negroes but is uncommon in Caucasians. A variant of TF\*D, TF\*D-CHI, appears at about the same frequency in Chinese as does TF\*D in Negroes. Isoelectric focusing is becoming the method of choice for GC and TF typing, so as to be able to discriminate GC\*1 and TF\*C subtypes. The results of a few subtyping studies are given in Tables 2 and 3, respectively. Additional data on these subtypes in U.S. populations would be highly desirable, and will doubtless be forthcoming from future surveys.

Table 4 gives the phenotypic proportions for relatively common hemoglobin  $\beta$ -chain variants, and Table 5 provides gene frequencies for those data for which meaningful calculations could be carried out. Although common hemoglobin variants have been employed as genetic markers in some laboratories using electrophoresis for many years [20-24], sometimes in conjunction with another established system [25], the application of isoelectric focusing procedures to their separation and identification is growing and may offer advantages in resolution [26-28].

It may be useful in certain circumstances to have frequency estimates for larger and presumably better randomized samples of the population at large than would result from local population studies, as we have discussed elsewhere [29,30]. Computation of WMP for systems in which a number of different studies have been done and in which fairly large numbers of people have been typed provides a possible approach to obtaining such an estimate. Populations are defined by the size of the effective interbreeding gene pool rather than by

TABLE 1—Genotypic and phenotypic frequencies of *haptoglobin α-chain (HPA) types in U.S. populations.*

Population	Total	Frequency—Number (Percent)					Gene Frequency, HPA #1	Note <sup>a</sup>	Reference
		1	2-1	2-1M	2	0			
				CAUCASIAN					
Ann Arbor, MI	68	9(13.2)	40(58.8)	...	19(27.9)	...	0.4265	...	T1
Michigan and Illinois	161	23(14.3)	77(47.8)	...	61(37.9)	...	0.3820	...	T2
Seattle, WA	409	54(13.2)	206(50.4)	...	149(36.4)	...	0.3839	1	T3
Southeastern GA	145	27(18.6)	64(44.1)	...	52(35.9)	2(1.4)	0.4126	...	T4
Maryland	192	24(12.5)	85(44.3)	...	83(43.2)	...	0.3464	...	T5
Chicago, IL	101	18(17.8)	51(50.5)	...	32(31.7)	...	0.4307	...	T6
Tecumseh, MI	7 655	1307(17.1)	3734(48.8)	11(0.1)	2575(33.6)	22(0.3)	0.4165	2	T7
Orange County, CA	185	(16.0)	(48.0)	...	(36.0)	...	...	3,4	T8
Bexar County, TX	200	(17.0)	(50.0)	...	(30.0)	...	...	3	T9
Pittsburgh and Allegheny County, PA	1 263	185(14.6)	555(43.9)	26(2.1)	495(39.2)	2(0.2)	0.3771	...	T10
California	274	(14.6)	(49.3)	...	(36.1)	...	...	3	T11
Philadelphia, PA, including part of NJ	212	33(15.6)	104(49.1)	...	75(35.4)	...	0.4009	5	T12
Detroit, MI	502	69(13.7)	213(42.4)	...	220(43.8)	...	0.3496	6	T13
Miami/Dade County, FL	366	58(15.8)	161(44.0)	...	141(38.5)	6(1.6)	0.3847	6	T15
Los Angeles, CA	311	62(19.9)	149(47.9)	...	100(32.2)	...	0.4389	6	T16
CA, TX, HI, and Mexico City	860	154(17.9)	418(48.6)	...	288(33.5)	...	0.4221	...	T17
U.S. national sample	5 735	(16.2)	(46.8)	...	(32.8)	...	...	3,7	T18
Los Angeles County, CA	393	63(16.0)	196(49.9)	...	134(34.1)	...	0.4097	...	T19
North Carolina	395	64(16.2)	199(50.4)	...	132(33.4)	...	0.4139	...	T20
				TOTAL CAUCASIAN					
Numerical total	13 033	2150(16.5)	6252(48.0)	37(0.3)	4556(35.0)	32(0.2)	0.4072	...	...
WMP	...	16.4	47.7	0.2	34.3	0.2	...	...	...
WSDP	...	1.194	1.953	0.497	2.580	0.268	...	...	...
				NEGRO					
Seattle, WA	406	(26.4)	(38.2)	(9.8)	(21.4)	(4.2)	...	3	T21
Ann Arbor, MI	48	17(35.4)	17(35.4)	...	9(18.7)	5(10.4)	0.5930	...	T1
Seattle, WA, and Cleveland, OH	178	45(25.3)	70(39.3)	27(15.2)	31(17.4)	5(2.8)	0.5405	...	T22

TABLE 1—(Continued).

Population	Total	Frequency—Number (Percent)				Gene Frequency, HPA* <i>f</i>	Note <sup>a</sup>	Reference
		1	2-1	2-1M	2			
New York, NY	100	(40.0)	(39.0)	...	(21.1)	...	3	T23
Seattle, WA	1 657	472(28.5)	641(38.7)	181(10.9)	307(18.5)	56(3.4)	8	T3
Southeastern GA	167	48(28.7)	61(36.5)	13(7.8)	42(25.1)	3(1.8)	...	T4
Chicago, IL	101	30(29.7)	47(46.5)	...	24(23.8)	...	...	T6
Bexar County, TX	200	(35.0)	(49.0)	...	(17.0)	...	3	T9
Pittsburgh and Allegheny County, PA	721	206(28.6)	293(40.6)	66(9.2)	148(20.5)	8(1.1)	...	T10
California	124	(33.9)	(41.9)	...	(21.0)	...	3,9	T11
Philadelphia, PA, including part of NJ	164	56(34.1)	73(44.5)	...	32(19.5)	3(1.8)	5	T12
Detroit, MI	504	142(28.2)	204(40.5)	59(11.7)	84(16.7)	11(2.2)	6,10 <sup>b</sup>	T13
Miami/Dade County, FL	346	120(34.7)	164(47.4)	...	40(11.6)	22(6.4)	6	T15
Los Angeles, CA	130	47(36.2)	58(44.6)	...	25(19.2)	...	6	T16
CA, TX, HI, and Mexico City	463	134(29.2)	236(51.0)	...	89(19.2)	...	11	T17
U.S. national sample	999	(32.2)	(39.4)	...	(13.3)	...	3,12	T18
Los Angeles County, CA	153	51(33.3)	77(50.3)	...	25(16.3)	...	...	T19
North Carolina	317	91(28.7)	142(44.8)	16(5.0)	68(21.5)	...	...	T20
TOTAL NEGRO								
Numerical total	4 949	1460(29.5)	2083(42.1)	362(7.3)	924(18.7)	113(2.3)	...	...
WMP	...	30.1	41.6	5.9	18.1	1.9	...	...
WSDP	...	2,910	4,134	5,255	3,138	1,963	...	...
HISPANIC								
Bexar County, TX	200	(25.0)	(51.0)	...	(24.0)	...	3	T9
California	161	(21.7)	(55.9)	...	(22.4)	...	3,13	T11
Miami/Dade County, FL	360	75(20.8)	179(49.7)	...	97(26.9)	9(2.5)	6	T15
Los Angeles, CA	145	43(29.7)	67(46.2)	...	35(24.1)	...	6	T16
CA, TX, HI, and Mexico City	775	218(28.1)	393(50.7)	...	163(21.0)	...	14,15	T17
Los Angeles County, CA	226	64(28.3)	103(45.6)	...	59(26.1)	...	...	T19

	1	506	400(26.6)	742(49.3)	TOTAL HISPANIC	354(23.5)	9(0.6)	0.5150	...
Numerical total	...	...	26.0	50.0	...	23.5	0.5	...	...
WMP	...	...	3.228	2.612	...	2.408	0.986	...	...
WSDP	...	...	...	...	...	...	...	...	...
					ORIENTAL AND ASIAN				
U.S. Japanese	23	2(8.7)		10(43.5)	...	11(47.8)	...	...	T23
New York, NY, Chinese	118	(14.7)		(38.3)	...	(47.0)	(0.03)	...	T24
Seattle, WA, Oriental	494	34(6.9)		190(38.5)	...	270(54.7)	...	0.2611	T3
New York, NY, Chinese	113	16(14.2)		45(39.8)	...	52(46.0)	...	0.3407	T25
California and Hawaii Asian	376	(7.7)		(37.8)	...	(53.5)	...	...	T11
CA, TX, HI, and									
Mexico City Asian	1	148(13.4)		444(40.2)	...	512(46.3)	...	0.3348	19
Los Angeles									
County, CA, Asian	12	2(16.7)		1(8.3)	...	9(75.0)	...	...	T19

\*NOTES:

1. HPA\*1 subtypes determined for 66 Type 1 and 2-1; there were 4(6.1) 1S-1, 5(7.6) 1S-1F, 36(54.5) 2-1S, and 21(31.8) 2-1F.
  2. Four were HP Ca and two were HP Johnson.
  3. Distributions given in percentages; data not used in calculating numerical totals.
  4. Approximately 15% of the population was Hispanic on the basis of surname.
  5. Identical twin study; data for one member of each twin pair tabulated and used in calculations.
  6. And see Shafer (Ref T14).
  7. 1.9% were "other."
  8. HPA\*1 subtypes determined for 222 type 1 and 2-1; there were 26(11.7) 1S-1S, 20(9.0) 1F-1F, 49(22.1) 1S-1F, 61(27.5) 2-1S, 55(24.8) 2-1F, 6(2.7) 2M-1S, and 5(2.3) 2M-1F.
  9. 3.2% were "rare."
  10. Four were "rare."
  11. Three were "rare."
  12. 12.1% were "other."
  13. "Chicano/Amerindian."
  14. One was "rare."
  15. Primarily "Mexican."
  16. HPA\*1 subtypes determined for 80 type 1 and 2-1; there were 14(17.5) 1S-1S, 1(1.3) 1S-1F, 64(80.0) 2-1S, and 1(1.3) 2-1F.
  17. HPA\*1 subtypes determined; all 16 type 1 were 1S-1S and all 45 type 2-1 were 2-1S.
  18. 1.1% were "rare."
  19. One was "rare."
- <sup>b</sup>χ<sup>2</sup> > 3.841; 0.01 < P < 0.05.  
<sup>c</sup>χ<sup>2</sup> > 6.635; P < 0.01.

TABLE 2—Genotypic and phenotypic frequencies of group specific component (GC; vitamin D binding protein) types in U.S. populations.

Population	Total	Frequency—Number (Percent)		Gene Frequency GC%	Note <sup>a</sup>	Reference
		1	2			
		CAUCASIAN				
New York, NY	122	48(55.8)	32(37.2)	0.7172	1	T26
New York, NY	86	147(50.3)	114(39.0)	0.7442	...	T27
Southeastern GA	292	234(57.5)	146(35.9)	0.6986	...	T28
Boston, MA	407	391(51.1)	313(41.0)	0.7543	...	T29
Tecumseh, MI	7 658	(50.2)	(41.0)	0.7155	...	T7
California	4 488			...	2,3	T11
Philadelphia, PA, including part of NJ	201	96(47.8)	93(46.3)	0.7090	4	T12
Detroit, MI	503	242(48.1)	213(42.3)	0.6928	5	T13
Miami/Dade County, FL	365	168(46.0)	160(43.8)	0.6795	5	T15
Los Angeles, CA	109	59(54.1)	47(43.1)	0.7569	5	T16
Southeastern PA	110	59(53.6)	40(36.4)	0.7182	6	T30
CA, TX, HI, and Mexico City	1 050	537(51.1)	429(40.9)	0.7157	7	T17
U.S. national sample	5 735	(51.9)	(38.5)	...	2	T18
Los Angeles County, CA	183	99(54.1)	78(42.6)	0.7541	<sup>b</sup>	T19
Minnesota	7 247	3742(51.6)	2963(40.9)	0.7208	8	T31
Southeastern MO	372	(55.1)	(39.3)	...	2	T32
		TOTAL CAUCASIAN				
Numerical total	18 211	9341(51.3)	7454(40.9)	0.7176	...	...
WMP	...	51.3	40.4	...	...	...
WSDP	...	1.350	1.365	...	...	...
		NEGRO				
New York, NY	144	...	...	0.8924	1	T26
New York, NY	120	98(81.7)	19(15.8)	0.8958	...	T27
Southeastern GA	231	192(83.1)	38(16.5)	0.9134	...	T28
California	832	(74.5)	(21.0)	...	2,9	T11
Philadelphia, PA, including part of NJ	159	122(76.7)	33(20.8)	0.8711	4	T12

Detroit, MI	504	366(72.6)	112(22.2)	40(8)	0.8562	5,10 <sup>c</sup>	T13
Miami/Dade County, FL	339	263(77.6)	64(18.9)	6(1.8)	0.8702	5,11	T15
Los Angeles, CA	43	34(79.1)	7(16.3)	2(4.7)	0.8721	5	T16
Southeastern GA	219	185(84.5)	26(11.9)	4(1.8)	0.9041	12 <sup>b</sup>	T30
Philadelphia, PA	273	198(72.5)	61(22.3)	3(1.1)	0.8370	13	T30
CA, TX, HI, and Mexico City	867	638(73.6)	183(21.1)	18(2.1)	0.8414	14	T17
U.S. national sample	999	(75.5)	(18.6)	(3.0)	...	2	T18
Los Angeles County, CA	75	59(78.7)	15(20.0)	1(1.3)	0.8867	...	T19
Minnesota	540	403(74.6)	104(19.3)	3(0.6)	0.8426	15	T31
Southeastern MO	69	(75.4)	(24.6)	(0.0)	...	2	T32
TOTAL NEGRO							
Numerical total	3 370	2558(75.9)	662(19.6)	49(1.5)	0.8573	...	...
WMP	...	75.6	19.7	1.8	...	...	...
WSDP	...	3.066	2.377	0.891	...	...	...
HISPANIC							
California	1 417	(59.1)	(35.3)	(5.2)	...	2,16	T11
Miami/Dade County, FL	360	207(57.5)	126(35.0)	24(6.7)	0.7500	5,17	T15
Los Angeles, CA	102	83(81.4)	17(16.7)	2(2.0)	0.8971	...	T16
CA, TX, HI, and Mexico City	1 908	1160(60.8)	655(34.3)	80(4.2)	0.7796	18	T17
Los Angeles County, CA	136	87(64.0)	38(27.9)	11(8.1)	0.7794	<sup>b</sup>	T19
TOTAL HISPANIC							
Numerical total	2 506	1537(61.3)	836(33.4)	117(4.7)	0.7801	...	...
WMP	...	60.5	34.1	4.9	...	...	...
WSDP	...	3.650	3.136	1.067	...	...	...
ASIAN AND ORIENTAL							
New York, NY, Chinese	117	...	...	...	0.7693	1	T26
California and Hawaii Asians	3 043	(50.4)	(39.2)	(6.4)	...	2	T11
CA, TX, HI, and Mexico City	1 566	780(49.8)	590(37.7)	118(7.5)	0.706	19 <sup>c</sup>	T17
Chinese and Japanese							

<sup>a</sup>NOTES:

1. Only gene frequencies and total number of people typed reported.
2. Distributions given in percentages; data not used in calculating numerical totals. (*footnotes continued*)



3. 0.4% were "rare."
  4. Identical twin study; data for one member of each twin pair tabulated and used in calculations.
  5. And see Schaler (Ref *T14*).
  6. GC\*1 subtypes determined; subtypes and rare types: 3(2.7) 1F, 19(17.3) 1F-1S, 37(33.6) 1S, 8(7.3) 2-1F, 32(29.0) 2-1S, and 11(10.0) 2.
  7. Ten were "rare."
  8. GC\*1 subtypes determined: 175(2.4) 1F, 1,266(17.5) 1F-1S, 2,301(31.8) 1S, 633(8.7) 2-1F, 2,330(32.2) 2-1S, and 542(7.5) 2.
  9. 2.4% were "rare."
  10. 18(3.6) were 1-1A1, 3(0.6) were 2-1A1, and 1 was "rare."
  11. 6(1.8) were 1-1A1.
  12. GC\*1 subtypes determined; subtypes and rarer types: 141(64.4) 1F, 40(18.3) 1F-1S, 4(1.8) 1S, 21(9.6) 2-1F, 5(2.3) 2-1S, 4(1.8) 2, 3(1.4) 1F-1A1, and 1(0.5) 2-1A1.
  13. GC\*1 subtypes determined; subtypes and rarer types: 130(47.6) 1F, 58(21.2) 1F-1S, 10(3.7) 1S, 51(18.7) 2-1F, 10(3.7) 2-1S, 3(1.1) 2, 6(2.2) 1F-1A1, 3(1.1) 1S-1A1, and 2(0.7) 2-1A1.
  14. 28 were "rare."
  15. GC\*1 subtypes determined; subtypes and rarer types: 244(45.2) 1F, 139(25.7) 1F-1S, 20(3.7) 1S, 89(16.5) 2-1F, 15(2.8) 2-1S, 3(0.6) 2, 7(1.3) 1F-1A1, 4(0.7) 1S-1A1, 2(0.4) 2-1A1, 9(1.7) 1F-1C10, 3(0.6) 1S-1C10, 3(0.6) 2-1C10, 1(0.2) 1A1, and 1(0.2) 1A1-1C10.
  16. "Chicano/Amerindian"; 0.5% were "rare."
  17. Two were 1-1A1 and 1 was 2-1A1.
  18. Primarily "Mexican"; 13 were "rare."
  19. Seventy eight were "rare."
- $b_{\chi^2} > 3.841$ ;  $0.01 < P < 0.05$ .
- \*Original author gene frequency value.

TABLE 3—Genotypic and phenotypic frequencies of transferrin (TF) types in U.S. populations.

Population	Frequency—Number (%)				Gene Frequency TF*C	Note <sup>a</sup>	Reference
	Total	C	BC	CD			
			CAUCASIAN				
Southeastern GA	107	103(96.3)	1(0.9)	2(1.9)	0.9766	1	T4
Southeastern U.S.	2 221	2194(98.8)	17(0.8)	10(0.5)	0.9939	2	T33
Chicago, IL	101	101(100.0)	...	...	1.0000	...	T6
Tecumseh, MI	7 654	7560(98.8)	83(1.1)	11(0.5)	0.9939	...	T7
Philadelphia, PA, including part of NJ	196	193(98.5)	2(1.0)	1(0.5)	0.9924	3	T12
CA, TX, HI, and Mexico City	801	786(98.1)	...	...	...	4	T17
U.S. national sample	5 735	(96.1)	(1.3)	(0.5)	...	5	T18
Philadelphia, PA	149	146(98.0)	2(1.3)	1(0.7)	0.9899	6	T34
Minnesota	392	392(100.0)	...	...	1.0000	7	T35
Minnesota	947	...	...	...	...	8	T36, T37
Baltimore, MD	209	205(89.1)	4(1.9)	...	0.9904	9	T38
			TOTAL CAUCASIAN				
Numerical total	4 176	4120(98.7)	26(0.6)	14(0.3)	0.9914	...	...
WMP	...	97.9	1.0	0.3	...	...	...
WSDP	...	1.275	0.349	0.226	...	...	...
			NEGRO				
New York, NY	99	89(89.9)	...	9(9.1)	0.9444	10	T24
Southeastern GA	133	120(90.2)	...	13(9.8)	0.9511	...	T4
Southeastern U.S.	418	399(95.5)	...	19(4.5)	0.9773	...	T33
Chicago, IL	101	93(92.1)	...	8(7.9)	0.9604	...	T6
Philadelphia, PA, including part of NJ	164	151(92.1)	...	13(7.9)	0.9604	3	T12
CA, TX, HI, and Mexico City	502	467(93.0)	...	...	...	11	T17
U.S. national sample	999	(90.8)	(0.2)	(5.8)	...	5	T18
Philadelphia, PA	166	151(91.0)	...	15(9.0)	0.9548	12	T34
Minnesota	194	182(93.8)	...	12(6.2)	0.9691	13	T35
Minnesota	194	...	...	...	...	14	T36, T37
Baltimore, MD	183	170(92.9)	1(0.6)	12(6.6)	0.9645	15	T38

TABLE 3—(Continued).

Population	Total	Frequency—Number (%)			Gene Frequency TF*C	Note <sup>a</sup>	Reference
		C	BC	CD			
Numerical total	1 960	1822(93.0)	1(0.1)	101(5.2)	0.9556	...	...
WMP	...	92.2	0.1	5.4	...	...	...
WSDP	...	1.707	0.147	2.803	...	...	...
		TOTAL NEGRO					
New York, NY—Chinese CA, TX, HI, and Mexico City—Hispanic	116	109(94.0)	...	7(6.0)	0.9698	16	T24
CA, TX, HI, and Mexico City—Asian	765	742(97.0)	...	...	...	17	T17
	1 295	1259(97.2)	...	...	...	18	T17
		OTHER					

<sup>a</sup>NOTES:

1. One was B1-2B2; the BC was B2C.
2. Two of the BC were B1C, and 15 were B2C.
3. Identical twin study; data for one member of each twin pair tabulated and used in calculations.
4. Fifteen were "rare."
5. Distributions given in percentages; data not used in calculating numerical totals.
6. TF\*C subtypes determined: 99(66.4) C1, 40(26.8) C2C1, 7(4.7) C2, 1(0.7) B2C1, 1(0.7) B2C2, and 1(0.7) C2D2.
7. TF\*C subtypes determined: 239(61.0) C1, 107(27.3) C2C1, 9(2.3) C2, 32(8.2) C3C1, 4(1.0) C3C2, and 1(0.3) C3.
8. Only gene frequencies and total number of people typed reported: TF\*C1 = 0.775, TF\*C2 = 0.163, TF\*C3 = 0.056, TF\*C6 = 0.001, TF\*B2 = 0.004, and TF\*B1-2 = 0.001.
9. TF\*C subtypes determined: 124(59.3) C1, 59(28.2) C2C1, 8(3.8) C2, 13(6.2) C3C1, 1(0.5) C3C2, 2(1.0) C1B2, 1(0.5) C1B1-2, and 1(0.5) C6B1-2.
10. One D1D1.
11. Thirty five were "rare."
12. TF\*C subtypes determined: 115(69.0) C1, 35(21.0) C2C1, 1(0.6) C2, 15(9.0) C1D1.
13. TF\*d subtypes determined: 137(70.6) C1, 38(19.6) C2C1, 4(2.1) C2, 3(1.6) C3C1, 11(5.7) C1D1, and 1(0.5) C2D1.
14. Only gene frequencies and total number of people typed reported; TF\*C1 = 0.842, TF\*C2 = 0.119, TF\*C3 = 0.008, TF\*C8 = 0.003, TF\*D1 = 0.028.
15. TF\*C subtypes determined: 134(73.2) C1, 29(15.9) C2C1, 3(1.6) C2, 4(2.2) C3C1, 1(0.5) C1B2, 10(5.5) C1D, 1(0.5) C1D2, and 1(0.5) C2D1.
16. The 7 CD were CD-CHI.
17. Primarily "Mexican"; 23 were "rare."
18. Thirty six were "rare."



TABLE 4—(Continued).

Population	Frequency—Number (Percent)										Reference	
	Total	A	AS	S	AD	AC	C	Note <sup>a</sup>				
Southern LA												
Children	564	479(84.9)	47(8.3)	18(3.2)	...	10(1.8)	...	5			T43	
Puerto Rico	602	561(93.2)	29(4.8)	2(0.3)	...	7(1.2)	1(0.2)	6			T50	
Philadelphia, PA												
Patients	1 000	897(89.7)	74(7.4)	3(0.3)	1(0.1)	23(2.3)	...	6			T51	
Baltimore, MD	400	...	...	...	4(1.0)	...	...	7			T52	
Memphis, TN												
Autopsy material	2 800	2 459(87.8)	254(9.1)	19(0.7)	1(0.04)	60(2.1)	...	8			T44	
Washington, DC												
Tuberculosis patients	310	282(91.0)	28(9.0)	...	...	...	...	...			T53	
St. Louis, MO												
Infants	449	359(80.0)	47(10.5)	...	2(0.4)	9(2.0)	...	9			T45	
Washington, DC												
Pregnant women study group	524	490(93.5)	25(4.8)	1(0.2)	...	8(1.5)	...	...			T54	
Control group	304	283(93.1)	11(3.6)	...	...	10(3.3)	...	...			T54	
Baltimore, MD												
Infants	900	784(87.1)	67(7.4)	...	...	26(2.9)	...	10			T46	
Maryland	681	625(91.8)	44(6.5)	...	...	12(1.8)	...	...			T55	
Southeastern GA	237	214(90.3)	19(8.0)	...	...	4(1.7)	...	...			T4	
Gainesville, FL												
Pregnant women	944	869(92.1)	65(6.9)	1(0.1)	...	9(1.0)	...	...			T56	
Mississippi	1 310	1 100(84.0)	114(8.7)	37(2.8)	14(1.1)	38(2.9)	7(0.5)	...			T47	
Southern LA												
Tuberculosis patients	220	211(95.9)	...	...	...	9(4.1)	...	...			T57	
Alabama	249 089	220 405(88.5)	21 423(8.6)	574(0.2)	...	6 074(2.4)	102(0.04)	11			T58	
California	1 025	(89.3)	(8.6)	...	...	(1.8)	...	1,12			T11	
Philadelphia, PA, including part of NJ	130	118(90.8)	11(8.5)	...	...	1(0.8)	...	13			T12	
Detroit, MI	504	452(89.7)	37(7.3)	...	...	14(2.8)	...	2,14			T13	
CA, TX, HI, and Mexico City	792	716(90.4)	54(6.8)	...	...	18(2.3)	...	15			T17	
Numerical total	265 220	234 707(88.5)	22 689(8.6)	670(0.3)	23(0.01)	6 421(2.4)	112(0.04)	...			...	
				TOTAL NEGRO								

WMP	...	88.5	8.5	0.3	0.0	2.4	0.0	...
WSDP	...	0.791	0.516	0.240	0.081	0.190	0.037	...
			HISPANIC					
Puerto Rico ("white")	1 487	1 486(99.9)	1(0.1)	...	...	...	...	T50
California	1 596	(99.6)	(0.1)	...	...	(0.2)	...	T11
CA, TX, HI, and Mexico City	1 569	1 561(99.5)	3(0.2)	...	...	3(0.2)	...	T17
			TOTAL HISPANIC					
Numerical total	3 056	3 047(99.7)	4(0.1)	...	...	3(0.1)	...	...
WMP	...	99.7	0.1	...	...	0.1	...	...
WSDP	...	0.177	0.052	...	...	0.091	...	...
			ASIAN					
California	3 053	(99.9)	...	...	...	...	...	T11
CA, TX, HI, and Mexico City	1 451	1 448(99.8)	1(0.1)	...	...	...	...	T17

NOTES:

1. Distributions given in percentages; data not used in calculating numerical totals.
2. And see Shaler, 1978 (Ref T14).
3. One was HBB SC.
4. Four were "other."
5. Ten were HBB SC.
6. Two were HBB SC.
7. An unspecified number of HBB AS were further tested to distinguish HBB AD; data not used in calculations.
8. Four were HBB SC; 3 were "other."
9. Thirty two were "other."
10. Twenty three were "other."
11. Of the HBB S, 149 were HBB\*S/HBB\*TH and 73 were HBB\*S/HBB\*High F; 329 were HBB SC, 11 were HBB S/Other, 7 were HBB C/Other, and 164 were rarer variants.
12. 0.4% were "rare."
13. Identical twin study; data for one member of each twin pair tabulated and used in calculations.
14. One was "rare."
15. Four were "rare."
16. Chicano/Amerindian; 0.1% were "rare."
17. Two were "rare."
18. 0.1% were "rare."

TABLE 5—Gene frequencies for common hemoglobin  $\beta$ -chain (*HBB*) variant genes in U.S. populations.

Population	Total Number Typed	Gene Frequencies				Reference to Population Study
		<i>HBB</i> *A	<i>HBB</i> *6V	<i>HBB</i> *6K	<i>HBB</i> *121Q	
CAUCASIAN						
Baltimore, MD	500	1.0000	...	...	...	T39
Houston, TX	350	1.0000	...	...	...	T41
Durham, NC	734	0.9986	0.0007	...	0.0007	T42
Southern LA						
Children	140	0.9964	0.0036	...	...	T43
Memphis, TN						
Autopsy material	1 250	1.0000	...	...	...	T44
St. Louis, MO						
Infants	90	1.0000	...	...	...	T45
Baltimore, MD						
Infants	180	1.0000	...	...	...	T46
Mississippi	1 045	0.9995	0.0005	...	...	T47
Detroit, MI	503	1.0000	...	...	...	T13
CA, TX, HI, and Mexico City	1 040	1.0000	...	...	...	T17
North Carolina	365	1.0000	...	...	...	T20
Total Caucasian	6 197	0.9996	0.0003	...	0.0001	...
NEGRO						
Baltimore, MD	500	0.9430	0.0460	0.0090	...	T39
St. Louis, MO	1 020	0.9392	0.0461	0.0128	0.0020	T48
Galveston, TX						
Patients	1 550	0.9450	0.0481	0.0119	...	T49
Houston, TX	400	0.9300	0.0575	0.0100	...	T41
Durham, NC	390	0.9269	0.0449	0.0167	0.0013	T42
Southern LA						
Children	564	0.8998	0.0825	0.0177	...	T43
Puerto Rico	602	0.9618	0.0291	0.0091	...	T50
Philadelphia, PA						
Patients	1 000	0.9455	0.0400	0.0110	0.0005	T51
Memphis, TN						
Autopsy material	2 800	0.9345	0.0529	0.0114	0.0002	T44
Washington, DC						
Tuberculosis patients	310	0.9548	0.0452	...	...	T53
St. Louis, MO						
Infants	449	0.8641	0.0523	0.0100	0.0022	T45
Washington, DC						
Pregnant women study group	524	0.9666	0.0258	0.0076	...	T54
Control group	304	0.9655	0.0181	0.0165	...	T54
Baltimore, MD						
Infants	900	0.9228	0.0372	0.0144	...	T46
Maryland	681	0.9589	0.0323	0.0088	...	T55
Southeastern GA	237	0.9515	0.0401	0.0084	...	T4
Gainesville, FL						
Pregnant women	944	0.9598	0.0355	0.0048	...	T56
Mississippi	1 310	0.9031	0.0718	0.0199	0.0053	T47
Southern LA						
Tuberculosis patients	220	0.9796	...	0.0205	...	T57
Alabama	249 089	0.9400	0.0457	0.0133	...	T58
Philadelphia, PA						
including part of NJ	130	0.9539	0.0423	0.0039	...	T12
Detroit, MI	504	0.9474	0.0367	0.0139	...	T13
CA, TX, HI, and Mexico City	792	0.9495	0.0341	0.0114	...	T17
Total Negro	265 220	0.9399	0.0453	0.0125	0.00004	...

TABLE 5—(Continued).

Population	Total Number Typed	Gene Frequencies				Reference to Population Study
		HBB*A	HBB*6V	HBB*6K	HBB*121Q	
HISPANIC						
Puerto Rico (“white”)	1 487	0.9997	0.0003	...	...	T50
CA, TX, HI, and Mexico City	1 569	0.9968	0.0010	0.0010	...	T17
ASIAN						
CA, TX, HI, and Mexico City	1 451	0.9983	0.0003	...	...	T17

geographical boundaries, especially in a country like the United States where mobility is very high. In a previous paper [14], we compared the phenotypic proportions for the ABO, Rh, and Secretor systems obtained in a recent study of a relatively large, completely ascertained, randomly selected countrywide population of Caucasian and Negro teenagers [T18] with the corresponding WMP values from our calculations. The results suggested that the WMP values compared favorably with the values obtained in the U.S. study. Table 6 gives a similar comparison of WMP values (with the data from the U.S. study omitted from the computation) with the U.S. data for the HP, GC, and TF systems. If the U.S. data are regarded as “expected” and the WMP data as “observed” in a chi-square goodness of fit test, the corresponding *P* exceeds 0.05 in every case. The greatest  $\chi^2$  value was obtained for Negro populations in the HP system. A major reason for this result is that the U.S. study did not report HP 2–1M and HP 0 proportions separately. A significant fraction of the 12.1% “other” types in this population would be expected to be HP 2–1M or HP 0. The comparison given in

TABLE 6—Comparison of WMP and U.S. phenotypic frequency values<sup>a</sup> for the HP, GC, and TF systems.

System	Phenotype	Proportion, %			
		Caucasian		Negro	
		U.S.	WMP	U.S.	WMP
HP	1	16.2	16.5	32.2	29.7
	2–1	46.8	48.3	39.4	48.8
	2	32.8	34.9	13.3	18.9
	0	<sup>b</sup>	0.2	<sup>b</sup>	2.3
		$\chi^2 = 0.1880$		$\chi^2 = 4.7946$	
GC	1	51.9	51.1	75.5	75.6
	2–1	38.5	40.9	18.6	20.0
	2	7.4	7.8	3.0	1.5
		$\chi^2 = 0.1836$		$\chi^2 = 0.8555$	
TF	C	96.1	98.7	90.8	93.0
	CB	1.3	0.9	0.2	0.1
	CD	0.5	0.2	5.8	5.2
			$\chi^2 = 0.3734$		$\chi^2 = 0.1654$

<sup>a</sup>Ref T18.

<sup>b</sup>1.9% Caucasian and 12.1% Negro reported as “other.”



Table 6 and the previous similar one involving the blood groups [14] suggest that the WMP value may provide a reasonably good estimate of the phenotypic proportions based upon the population data which have been gathered.

#### Acknowledgments

We thank the following for kindly sharing the results of their unpublished population studies for inclusion in the data presented: W. C. Stuver of the Miami/Dade County Metro Police Department Crime Laboratory, Gary Siglar of the Los Angeles Police Department Crime Laboratory, Barry A. J. Fischer of the Los Angeles County Sheriff's Department Criminalistics Laboratory, and Dr. R. C. Briner of the Southeast Missouri Regional Crime Laboratory. In addition, we thank Holly Hammond, now of the Houston Police Department Crime Laboratory, and Matthew Abbott, now of the Maryland State Police Laboratory, for their assistance in modifying programs and management of the extensive data files.

#### Special Acknowledgments

We take particular pleasure in thanking Dr. Peter Shenkin, Department of Mathematics, John Jay College of Criminal Justice of the City University of New York, and Dr. James Uebelacker, Department of Mathematics, University of New Haven, for their many helpful discussions and suggestions.

#### References

- [1] Gaensslen, R. E. and Camp, F. R., "Forensic Serology: Parentage Testing," in *Forensic Sciences*, Vol. 2, C. H. Wecht, Ed., Matthew Bender, New York, 1986, Chap. 30 (originally as F. R. Camp and R. E. Gaensslen, "Forensic Serology: Paternity Testing (and Transfusion Reactions)," same title, 1981).
- [2] *Probability of Inclusion in Paternity Testing*, H. Silver, Ed., American Association of Blood Banks, Arlington, VA, 1982.
- [3] *Inclusion Probabilities in Parentage Testing*, R. H. Walker, Ed., American Association of Blood Banks, Arlington, VA, 1983.
- [4] Culliford, B. J., *The Examination and Typing of Bloodstains in the Crime Laboratory*, U.S. Government Printing Office, Washington, DC, 1971.
- [5] Metropolitan Police Forensic Science Laboratory, *Biology Methods Manual*, Commissioner of Police of the Metropolis, London, 1978.
- [6] Gaensslen, R. E. and Camp, F. R., "Forensic Serology: Analysis of Bloodstains and Body Fluid Stains," in *Forensic Sciences*, C. H. Wecht, Ed., Matthew Bender, New York, 1984, Vol. 2, Chapter 29.
- [7] Lee, H. C., "Identification and Grouping of Bloodstains," in *Forensic Science Handbook*, R. Saferstein, Ed., Prentice Hall, Englewood Cliffs, NJ, 1982, pp. 267-337.
- [8] Sensabaugh, G. F., "Isozymes in Forensic Science," in *Isozymes. Current Topics in Biological and Medical Research*, Vol. 6, M. C. Rattazzi, J. G. Scandalios, and G. S. Whitt, Eds., Alan R. Liss, New York, 1982, pp. 247-282.
- [9] Sensabaugh, G. F., "The Utilization of Polymorphic Enzymes in Forensic Science," in *Isozymes. Current Topics in Biological and Medical Research*, Vol. 11, M. C. Rattazzi, J. G. Scandalios, and G. S. Whitt, Eds., Alan R. Liss, New York, 1983, pp. 137-154.
- [10] Gaensslen, R. E., *Sourcebook in Forensic Serology, Immunology and Biochemistry*, U.S. Government Printing Office, Washington, DC, 1983.
- [11] Mourant, A. E., Kopeć, A. C., and Domaniewska-Sobczak, K., *The Distribution of the Human Blood Groups and Other Polymorphisms*, Oxford University Press, London, 1976.
- [12] Walter, H. and Steegmüller, H., "Studies on the Geographical and Racial Distribution of the Hp and Gc Polymorphisms," *Human Heredity*, Vol. 19, 1969, pp. 209-221.
- [13] Livingstone, F. B., *Abnormal Hemoglobins in Human Populations*, Aldine, Chicago, 1967.
- [14] Gaensslen, R. E., Bell, S. E., and Lee, H. C., "Distributions of Genetic Markers in United States Populations: I. Blood Group and Secretor Systems," *Journal of Forensic Sciences*, Vol. 32, No. 4, July 1987, pp. 1016-1058.

- [15] Gaensslen, R. E., Bell, S. E., and Lee, H. C., "Distributions of Genetic Markers in United States Populations: II. Isoenzyme Systems," *Journal of Forensic Sciences*, Vol. 32, No. 5, Sept. 1987, pp. 1348-1381.
- [16] Giblett, E. R., *Genetic Markers in Human Blood*, Blackwell Scientific Publications, Oxford & Edinburgh, U.K., 1969.
- [17] Giblett, E. R. and Steinberg, A. G., "The Inheritance of Serum Haptoglobin Types in American Negroes: Evidence for a Third Allele  $Hp^{2M}$ ," *American Journal of Human Genetics*, Vol. 12, 1960, pp. 160-169.
- [18] Shows, T. B., Alper, C. A., Bootsma, D., Dorf, M., Douglas, T., et al., "International System for Human Gene Nomenclature (1979). ISGN (1979)," *Cytogenetics and Cell Genetics*, Vol. 25, 1979, pp. 96-116.
- [19] Constans, J. and Cleve, H., "Group-specific Component. Report on First International Workshop," *Human Genetics*, Vol. 48, 1979, pp. 143-149.
- [20] Pollack, O. J., Barug, E. R., and Chubaty, W. G., "Toxicologic Study of Chemically Altered Hemoglobins and Forensic Study of Blood Stains by Paper Electrophoresis," *Journal of Forensic Medicine*, Vol. 5, 1958, pp. 200-207.
- [21] Huntsman, R. G. and Lehmann, H., "The Detection of Abnormal and Foetal Haemoglobins in Blood Stains," *Medicine Science and the Law*, Vol. 3, 1962, pp. 59-64.
- [22] Culliford, B. J., "The Identification of Abnormal Haemoglobins in Bloodstains," *Journal of the Forensic Science Society*, Vol. 4, 1964, pp. 155-157.
- [23] Wraxall, B. G. D., "The Identification of Foetal Haemoglobin in Bloodstains," *Journal of the Forensic Science Society*, Vol. 12, 1972, pp. 457-458.
- [24] Wiggins, K. G., "The Identification of Five Hemoglobin Variants in Bloodstains," *Journal of the Forensic Science Society*, Vol. 18, 1978, pp. 57-60.
- [25] Divall, G. B. and Greenhalgh, M., "The Screening of Blood Samples for Haemoglobin Variation in Conjunction with Glyoxalase Typing," *Journal of the Forensic Science Society*, Vol. 23, 1983, pp. 49-51.
- [26] Burdett, P. E. and Whitehead, P. H., "The Separation of the Phenotypes of Phosphoglucomutase, Erythrocyte Acid Phosphatase, and Some Haemoglobin Variants by Isoelectric Focusing," *Analytical Biochemistry*, Vol. 77, 1977, pp. 419-428.
- [27] Bassett, P., Beuzard, V., Garel, M. C., and Rosa, J., "Isoelectric Focusing of Human Hemoglobin: Its Application to Screening, to the Characterization of 70 Variants, and to the Study of Modified Fractions of Normal Hemoglobins," *Blood*, Vol. 51, 1978, pp. 971-982.
- [28] Bonte, W., Jursch, R., and Straube, J., "Identification of Hemoglobin Derivatives by Means of Electrofocusing," *Forensic Science International*, Vol. 17, 1981, pp. 45-49.
- [29] Gaensslen, R. E., Lee, H. C., Ehart, S., Abbott, M., and Hammond, H., "Use and Interpretation of Phenotypic Frequencies for Genetic Markers in Populations in Forensic Serology," in *Proceedings*, International Symposium on the Forensic Applications of Electrophoresis, FBI Academy, Quantico, VA, 1984.
- [30] Gaensslen, R. E., "When Blood is Their Argument: Use and Interpretation of Population Genetic Marker Frequency Data in Forensic Serology," *FBI Crime Laboratory Digest*, Vol. 12, No. 4, Oct. 1985, pp. 75-81.

### Table References

- [T1] Sutton, H. E., Neel, J. V., Livingstone, F. B., Binson, G., Kunstadter, P., and Trombley, L. E., "The Frequencies of Haptoglobin Types in Five Populations," *Annals of Human Genetics*, Vol. 23, 1959, pp. 175-183.
- [T2] Bayani-Sioson, P. S., Louch, J., Sutton, H. E., Neel, J. V., Horne, S. L., and Gershowitz, H., "Quantitative Studies on the Haptoglobin of Apparently Healthy Adult Male Twins," *American Journal of Human Genetics*, Vol. 14, 1962, pp. 210-219.
- [T3] Giblett, E. R. and Brooks, L. E., "Haptoglobin Types: Haptoglobin Subtypes in Three Racial Groups," *Nature*, Vol. 197, 1963, pp. 576-577.
- [T4] Cooper, A. J., Blumberg, B. S., Workman, P. L., and McDonough, J. R., "Biochemical Polymorphic Traits in a U.S. White and Negro Population," *American Journal of Human Genetics*, Vol. 15, 1963, pp. 420-428.
- [T5] Queen, K. G. and Peacock, A. C., "Haptoglobin Type Determination Using Noncarcinogenic Reagents. Results on 192 Normal Subjects in Maryland," *Clinica Chimica Acta*, Vol. 13, 1966, pp. 47-51.
- [T6] Shih, L.-Y. and Hsia, D. Y.-Y., "The Distribution of Genetic Polymorphisms Among Chinese in Taiwan," *Human Heredity*, Vol. 19, 1969, pp. 227-233.
- [T7] Schreffler, D. C., Sing, C. F., Neel, J. V., Gershowitz, H., and Napier, J. A., "Studies on Ge-

- netic Selection in a Completely Ascertained Caucasian Population. I. Frequencies, Age and Sex Effects, and Phenotype Associations for 12 Blood Group Systems," *American Journal of Human Genetics*, Vol. 23, 1971, pp. 150-163.
- [T8] Fitzpatrick, F. A., Graves, M. H., and White, J. M., "The Distribution of EsD Phenotypes in a Selected California Population," *Forensic Serology News*, Vol. 2, No. 6, 1976.
- [T9] Ganaway, R. L. and Lux, P., "ABO, PGM and Hp Distribution in Three Ethnic Groups in Bexar County, Texas," *Forensic Serology News*, Vol. 4, No. 1, 1978.
- [T10] Hagins, A. M., Shaler, R. C., Mortimer, C. M., Stuver, W. C., and Neilson, D. M., "Population Frequencies of Forensically Important Genetic Markers: Phosphoglucomutase, Erythrocyte Acid Phosphatase, and Haptoglobin," *Journal of Forensic Sciences*, Vol. 23, No. 3, July 1978, pp. 563-569.
- [T11] Grunbaum, B. W., Selvin, S., Pace, N., and Black, D. M., "Frequency Distribution and Discrimination Probability of Twelve Protein Genetic Variants in Human Blood as Functions of Race, Sex and Age," *Journal of Forensic Sciences*, Vol. 23, No. 3, July 1978, pp. 577-587.
- [T12] Pakstis, A. J., Polesky, H. F., Scarr, S., and Katz, S. H., "Gene Frequency Estimates for Samples of Black and White Twins from the Philadelphia Metropolitan Area," *Human Genetics*, Vol. 43, 1978, pp. 159-177.
- [T13] Stolorow, M. D., Housel, D. L., Schaefer, J. R., Schoonover, J. L., Hauncher, J. D., Metzger, D. A., and Backos, G. B., "Blood Genetic Marker Study—Greater Detroit Metropolitan Area," paper presented at 31st Annual Meeting, American Academy of Forensic Sciences, Atlanta, GA, Feb. 1979.
- [T14] Shaler, R. C., *Forensic Implications of Genetic Population Data Collected in Different Geographical Regions*, Final Report, Report ATR-79(7910)-1, Contract J-LEAA-025-73 for Law Enforcement Assistance Administration, Eastern Technical Division, Aerospace Corp., 1978.
- [T15] Stuver, W. C., unpublished personal communication, 1979.
- [T16] Siglar, G., unpublished personal communication, 1979.
- [T17] Grunbaum, B. W., Selvin, S., Myhre, B. A., and Pace, N., "Distribution of Gene Frequencies and Discrimination Probabilities for 22 Human Blood Genetic Systems in Four Racial Groups," *Journal of Forensic Sciences*, Vol. 25, No. 2, April 1980, pp. 428-444.
- [T18] United States National Center for Health Statistics, *Selected Genetic Markers of Blood and Secretions for Youths, 12-17 Years of Age*, United States, DHEW Publ. (PHS) 80-1664, U.S. Department of Health, Education and Welfare, Public Health Service, Office of Health Research, Statistics and Technology, National Center for Health Statistics, Hyattsville, MD, 1980.
- [T19] Fischer, B., unpublished personal communication, 1982.
- [T20] Nelson, M. S., "A Computer-Assisted Population Frequency Study of 14 Polymorphic Blood Group Systems in North Carolina," *Journal of Forensic Sciences*, Vol. 29, No. 3, July 1984, pp. 762-773.
- [T21] Giblett, E. R., "Haptoglobin Types in American Negroes," *Nature*, Vol. 183, 1959, pp. 192-193.
- [T22] Giblett, E. R. and Steinberg, A. G., "The Inheritance of Serum Haptoglobin Types in American Negroes: Evidence for a Third Allele, Hp2m," *American Journal of Human Genetics*, Vol. 12, 1960, pp. 160-169.
- [T23] Harris, H., Robson, E. B., and Siniscalco, M., "Genetics of the Plasma Protein Variants," in *CIBA Foundation Symposium on Biochemistry of Human Genetics*, G. E. W. Wolstenholme and C. M. O'Connor, Eds., Little Brown, Boston, 1959, pp. 151-177.
- [T24] Parker, W. C. and Bearn, A. G., "Haptoglobin and Transferrin Variation in Humans and Primates: Two New Transferrins in Chinese and Japanese Populations," *Annals of Human Genetics*, Vol. 25, 1961, pp. 227-241.
- [T25] Shim, B.-S. and Bearn, A. G., "The Distribution of Haptoglobin Subtypes in Various Populations, Including Subtype Patterns in Some Nonhuman Primates," *American Journal of Human Genetics*, Vol. 16, 1964, pp. 477-483.
- [T26] Cleve, H. and Bearn, A. G., "Studies on the 'Group Specific Component' of Human Serum. Gene Frequencies in Several Populations," *American Journal of Human Genetics*, Vol. 13, 1961, pp. 372-378.
- [T27] Cleve, H. and Bearn, A. G., "Inherited Variations in Human Serum Proteins: Studies on the Group-Specific Component," *Annals of the New York Academy of Sciences*, Vol. 94, 1961, pp. 218-224.
- [T28] Blumberg, B. S., Workman, P. L., and Hirschfeld, J., "Gamma-Globulin, Group Specific and Lipoprotein Groups in a U.S. White and Negro Population," *Nature*, Vol. 202, 1964, pp. 561-563.
- [T29] Murray, R. F. and Robison, J. C., "Observations on the Distribution of Group Specific Component (Gc) Types in Subjects Who Have Had Rheumatoid Arthritis," *Acta Genetica et Statistica Medica*, Vol. 18, 1968, pp. 399-405.

- [T30] Kueppers, F. and Harpel, B., "Group-Specific Component (Gc) 'Subtypes' of Gc 1 by Isoelectric Focusing in U.S. Blacks and Whites," *Human Heredity*, Vol. 29, 1979, pp. 242-249.
- [T31] Dykes, D. D., Crawford, M. H., and Polesky, H. F., "Population Distribution in North and Central America of PGM1 and Gc Subtypes as Determined by Isoelectric Focusing (IEF)," *American Journal of Physical Anthropology*, Vol. 62, 1983, pp. 137-145.
- [T32] Briner, R. C. and Compass, T., unpublished personal communication, 1985.
- [T33] Roop, W. E., Roop, B. L., and Putnam, F. W., "Transferrin Variants among Blood Donors," *Vox Sanguinis*, Vol. 14, 1968, pp. 255-257.
- [T34] Kueppers, F. and Harpel, B. M., "Transferrin C Subtypes in US Blacks and Whites," *Human Heredity*, Vol. 30, 1980, pp. 376-382.
- [T35] Dykes, D. and Polesky, H. F., "Transferrin (Tf) Subtyping on Agarose: A New Technique for Isoelectric Focusing," *Human Genetics*, Vol. 59, 1981, pp. 365-366.
- [T36] Dykes, D. D., DeFurio, C. M., and Polesky, H. F., "Transferrin (Tf) Subtypes in U.S. Amerindians, Whites and Blacks Using Thin-Layer Agarose Gels: Report on a New Variant, Tf<sup>CS</sup>," *Electrophoresis*, Vol. 3, 1982, pp. 162-164.
- [T37] Dykes, D. D., DeFurio, C. M., and Polesky, H. F., "Isoelectric Focusing for Transferrin (Tf) Subtypes in Parentage Testing," *American Journal of Clinical Pathology*, Vol. 79, 1983, pp. 725-727.
- [T38] Budowle, B., "Transferrin Subtypes Determined by Ultrathin-Layer Polyacrylamide Gel Isoelectric Focusing," *Electrophoresis*, Vol. 6, No. 2, Feb. 1985, pp. 97-99.
- [T39] Smith, E. W. and Conley, C. L., "Filter Paper Electrophoresis of Human Hemoglobins with Special Reference to the Incidence and Clinical Significance of Hemoglobin C," *Bulletin of the Johns Hopkins Hospital*, Vol. 93, 1953, pp. 94-106.
- [T40] Neel, J. V., "Implications of Some Recent Developments in Hematological and Serological Genetics," *American Journal of Human Genetics*, Vol. 6, 1954, pp. 208-223.
- [T41] Haynie, T. P., Dobson, H. L., and Hettig, R. A., "Molecular Diseases of Hemoglobin. I. Introduction and Incidence," *Annals of Internal Medicine*, Vol. 46, 1957, pp. 1031-1038.
- [T42] Chernoff, A. I. and Weichselbaum, T. E., "A Microhemolyzing Technic for Preparing Solutions of Hemoglobin for Paper Electrophoretic Analysis," *American Journal of Clinical Pathology*, Vol. 30, 1958, pp. 120-125.
- [T43] Moffitt, E. M. and McDowell, C. W., "The Incidence of Abnormal Hemoglobins by Paper Electrophoresis in Southern Louisiana," *Bulletin of the Tulane Medical Faculty*, Vol. 19, 1959, pp. 167-180.
- [T44] McCormick, W. F., "Abnormal Hemoglobins. I. Incidence in Memphis and Western Tennessee with Special Reference to Autopsy Material," *American Journal of Clinical Pathology*, Vol. 34, 1960, pp. 220-224.
- [T45] Minnich, V., Cordonnier, J. K., Williams, W. J., and Moore, C. V., "Alpha, Beta and Gamma Hemoglobin Polypeptide Chains During the Neonatal Period with a Description of a Fetal Form of Hemoglobin, D<sub>St. Louis</sub>," *Blood*, Vol. 19, 1962, pp. 137-167.
- [T46] Weatherall, D. J., "Abnormal Haemoglobins in the Neonatal Period and Their Relationship to Thalassaemia," *British Journal of Haematology*, Vol. 9, 1963, pp. 265-277.
- [T47] Thompson, R. B., Legan, S., and Odom, J., "A Survey of Hemoglobinopathies in Mississippi," *Journal of the Mississippi State Medical Association*, Vol. 5, 1964, pp. 461-465.
- [T48] Chernoff, A. I., "On the Presence of Hemoglobin D in the American Negro," *Blood*, Vol. 11, 1956, pp. 907-909.
- [T49] Schneider, R. G., "Incidence of Electrophoretically Distinct Abnormalities of Hemoglobin in 1550 Negro Hospital Patients," *American Journal of Clinical Pathology*, Vol. 26, 1956, pp. 1270-1276.
- [T50] Suarez, R. M., Buso, R., Meyer, L. M., and Olavarrieta, S. T., "Distribution of Abnormal Hemoglobins in Puerto Rico and Survival Studies of Red Blood Cells Using Cr<sup>51</sup>," *Blood*, Vol. 14, 1959, pp. 255-261.
- [T51] Myerson, R. M., Harrison, E., and Lohmuller, H. W., "Incidence and Significance of Abnormal Hemoglobins. Report of a Series of 1,000 Hospitalized Negro Veterans," *American Journal of Medicine*, Vol. 26, 1959, pp. 543-546.
- [T52] Marder, V. J. and Conley, C. L., "Electrophoresis of Hemoglobin on Agar Gels. Frequency of Hemoglobin D in a Negro Population," *Bulletin of the Johns Hopkins Hospital*, Vol. 105, 1959, pp. 77-88.
- [T53] Ryan, T. J., O'Connor, T. F., McCurdy, P. R., and Katz, S., "Sickle Cell Trait and Tuberculosis," *American Review of Respiratory Disease*, Vol. 81, 1960, pp. 546-549.
- [T54] Jenkins, M. E. and Clark, J. F. J., "Studies into Maternal Influence on the Well-Being of the Fetus and Newborn. I. The Distribution of Abnormal Hemoglobins among Pregnant Negro Women," *American Journal of Obstetrics and Gynecology*, Vol. 84, 1962, pp. 57-61.
- [T55] Boyer, S. H., Rucknagel, D. L., and Weatherall, D. J., "Further Evidence for Linkage Between

- the  $\beta$  and  $\delta$  Loci Governing Human Hemoglobin and the Population Dynamics of Linked Genes," *American Journal of Human Genetics*, Vol. 15, 1963, pp. 438-448.
- [T56] Cotter, J. and Prystowski, H., "Routine Hemoglobin Electrophoresis in Negro Gravidas," *Obstetrics and Gynecology*, Vol. 22, 1963, pp. 610-611.
- [T57] Coulter, W. W., "Incidence of Hemoglobin 'C' Trait in Tuberculosis Negroes," *Journal of the Louisiana State Medical Society*, Vol. 117, 1965, pp. 242-244.
- [T58] Schneider, R. G., Hightower, B., Hosty, T. S., Ryder, H., Tomlin, G., et al., "Abnormal Hemoglobins in a Quarter Million People," *Blood*, Vol. 48, 1976, pp. 629-637.

Address requests for reprints or additional information to  
R. E. Gaensslen, Ph.D.  
University of New Haven  
300 Orange Ave.  
West Haven, CT 06516